



## Mouse NFκB-Regulated cDNA Plate Array

Catalog Number AP-1101

(For Research Use Only)

### Introduction

NFκB is a ubiquitous transcription factor that plays a key role in cellular responses to stimuli such as stress, cytokines, free radicals, ultraviolet irradiation, oxidized LDL, and bacterial or viral antigens. Incorrect regulation of NFκB has been linked to cancer, inflammatory and autoimmune diseases, septic shock, viral infection, and improper immune development. When NFκB is activated, it is dissociated from its inhibitor IκB and moves from the cytoplasm to the nucleus, where it binds to target DNA elements and positively regulates the transcription of genes involved in immune and inflammatory responses, cell growth control, and apoptosis. Signosis developed a plate-based array for profiling 20+ mouse NFκB-regulated genes.

### Principle of the assay

Signosis' proprietary cDNA plate array is a plate-based hybridization profiling analysis for monitoring the expression of dozens of genes through reverse transcription of mRNA into cDNA. Like array analyses, total RNA is first reverse transcribed into cDNA in the presence of biotin-dUTP in the assay. Targeted genes are then specifically captured onto individual wells on a plate, instead of membranes, through a pre-coated gene-specific oligonucleotide. The captured cDNAs are further detected with streptavidin-HRP. The chemiluminescent signals are read with a plate reader. The expression level of genes is directly proportional to the chemiluminescent intensity.

### Materials provided with the kit

- A 96-well plate coated with 23 different capture oligos (RT)
- Mouse NFκB Reg. Primer Mix (-20 °C)
- Reverse transcription buffer mix (-20 °C)
- Reverse transcriptase RT (-20 °C)
- Streptavidin-HRP conjugate (4 °C)
- Plate hybridization buffer (RT)
- 5x Plate hybridization wash buffer (RT)
- Blocking buffer (RT)
- 5xDetection wash buffer (RT)
- Substrate A (4 °C)
- Substrate B (4 °C)
- Substrate dilution buffer (RT)

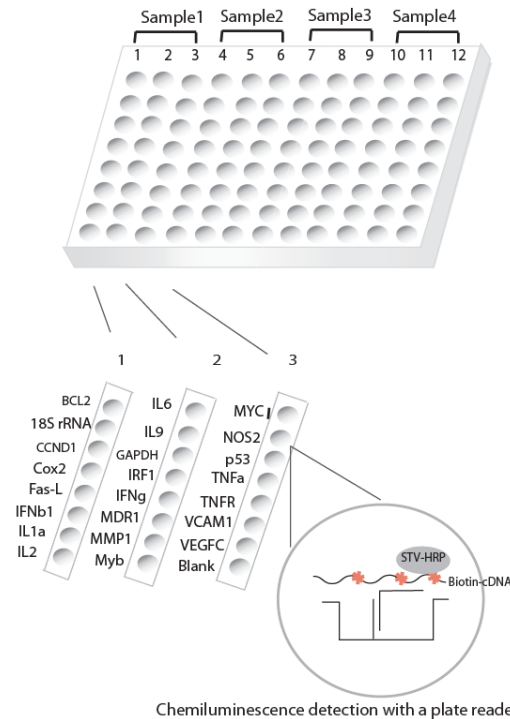


Diagram of Mouse NFκB-Regulated cDNA Plate Array

### Material required but not provided

PCR machine  
Incubator  
0.2ml PCR tube  
Luminometer plate reader  
ddH<sub>2</sub>O (RNAase free)

### Reagent preparation before starting experiment

- Dilute 30ml of 5x Plate hybridization wash buffer with 120 ml of dH<sub>2</sub>O before use.
- Dilute 40ml of 5x Detection wash buffer with 160 ml of dH<sub>2</sub>O before use.
- Warm up Plate hybridization buffer and 1x Plate hybridization wash buffer at 45 °C until no visible precipitate before use. Stir the solution with 10ml or 5ml pipette to facilitate the dissolving process.
- Dilute 500 times of streptavidin-HRP with blocking buffer before use at Step 3(4).

## Assay procedure

### 1. cDNA synthesis using PCR machine

**Note: Briefly spin tubes before opening**

(1) Sample preparation

X µl 1-10µg total RNA  
2 µl Mouse NFκB Reg. Primer Mix  
X µl ddH<sub>2</sub>O

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11µl

- (2) Incubate for 5 minutes at 65 °C, and chill on ice.  
(3) Add 8 µl Reverse transcription buffer mix and 1µl RT to each reaction tube, and incubate for 1 hour at 45 °C.  
(4) Heat the reaction to 98 °C for 5 minutes, and chill on ice.  
(5) The 20ul cDNA is synthesized and labeled with biotin and ready for hybridization on the plate.

### 2. Plate hybridization

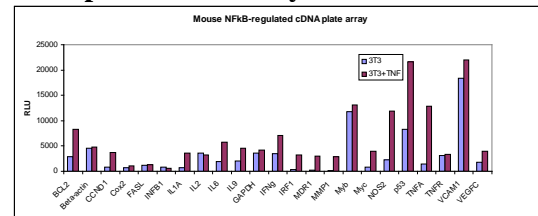
- (1) Remove the sealing film  
(2) Arrange the appropriate number of the wells of the plate based on your experiment. The whole plate is divided into 4 repeat sections, Column 1-3, 4-6, 7-9, 10-12 for 4 different samples.  
(3) Mix 20ul cDNA with 2.8ml pre-warmed Plate hybridization buffer, and dispense 95ul mixture to each well in a section **immediately**. A reagent reservoir can be used for dispensing cDNA mixture into the wells with a 8 multichannel pipette. Add 100ul Plate hybridization buffer without cDNA to the 'Blank' well.  
(4) Seal the wells with foil film securely and hybridize at 45 °C for overnight. Ensure the numbers and letters on the plate are clearly visible from under foil seal by pressing the foil down on every single experimental well. *Put an open container with water in the incubator to keep humidity and prevent evaporation from experimental wells.*

### 3. Plate detection

- (1) Removing the top foil sealing film with a blade to expose the experimental wells. Keep the unused well sealed for the future usage.  
(2) Invert the plate over an appropriate container and expel the contents forcibly, and wash the plate by adding 300µl of warmed 1x Plate hybridization wash buffer. Repeat the washing process two times for a total of three washes. Complete removal of liquid at each wash by firmly tapping the plate against clean paper towels.

- (3) Add 200µl of Blocking buffer incubate for 15 minutes at room temperature with gentle shaking.  
(4) Invert the plate over an appropriate container to remove blocking buffer. And add 100 µl of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.  
(5) Invert the plate over an appropriate container and expel the contents forcibly, and wash the plate with 200ul 1X Detection wash buffer for 5 min at room temperature with gently shaking. Complete removal of liquid at each wash by firmly tapping the plate against clean paper towels.  
(6) Repeat (5) for additional 2 time washes.  
(7) Freshly prepare the substrate solution  
For the whole plate:  
1ml Substrate A  
1ml Substrate B  
8ml Substrate dilution buffer  
(8) Add 95µl substrate solution to each well and incubate for 1minutes.  
(9) Place the plate in the luminometer, and read. Set integration time to 1 second with no filter position. For the best results, read the plate within 5-20 minutes.

### Example of Data Analysis



**Figure1: Mouse NFκB-regulated cDNA plate assay for analysis of 21 genes.**

NIH3T3 cells were treated with and without 20ng/ul TNF for 90 minutes respectively. Total RNAs were prepared. RNAs were subjected to NFκB regulated cDNA plate assay, and detected with chemiluminescence by a plate reader.

### Diagram of Mouse NFκB cDNA plate assay

	1	2	3	4	5	6	7	8	9	10	11	12
A	BCL2	IL6	Myc	BCL2	IL6	Myc	BCL2	IL6	Myc	BCL2	IL6	Myc
B	18S rRNA	IL9	NOS2	18S rRNA	IL9	NOS2	18S rRNA	IL9	NOS2	18S rRNA	IL9	NOS2
C	CCND1	GAPDH	p53	CCND1	GAPDH	p53	CCND1	GAPDH	p53	CCND1	GAPDH	p53
D	Cox2	IFNG	TNFA	Cox2	IFNG	TNFA	Cox2	IFNG	TNFA	Cox2	IFNG	TNFA
E	FASL	IRF1	TNFR	FASL	IRF1	TNFR	FASL	IRF1	TNFR	FASL	IRF1	TNFR
F	INFB1	MDR1(Abc	VCAM1	INFB1	MDR1(Abc	VCAM1	INFB1	MDR1(Abc	VCAM1	INFB1	MDR1(Abc	VCAM1
G	IL1A	MMP1	VEGFC	IL1A	MMP1	VEGFC	IL1A	MMP1	VEGFC	IL1A	MMP1	VEGFC
H	IL2	Myb	Blank	IL2	Myb	Blank	IL2	Myb	Blank	IL2	Myb	Blank