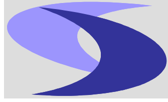

Signosis
BioSignal Capture

Mouse Cytokine ELISA Plate Array I (Chemiluminescence)

Catalog Number EA-4003 **(For Research Use Only)**

Introduction

Cytokines are signaling molecules that have critical roles in many biological processes such as cellular growth, differentiation, gene expression, migration, immunity and inflammation. Cytokines are secreted from cells and bound to cell-surface receptors, which initiate the activation of signal transduction pathways and mediate cell to cell communication. The malfunction of cytokines leads to many diseases, including arthritis, acute and chronic liver disease, inflammatory bowel disease, cardiac-related diseases and cancers. A group of cytokines commonly involves in one biological or disease process, therefore, the comprehensive analysis of the expression of multiple cytokines allows revealing the underneath mechanism of the disease state effectively. The Mouse Cytokine ELISA Plate Array I allows you to monitor the abundance of 23 mouse cytokines in a high-throughput manner. This assay is a fast and sensitive tool for quantitatively profiling the levels of multiple cytokines between samples simultaneously.

Principle of the assay

The 96-well white plate is divided into 4 sections, and each section has 3 columns for one sample. In each section, 23 of specific cytokine capture antibodies are coated on 23 wells respectively, and one well without coating any antibody is used as a blank well. The sample, such as cell culture supernatants, cell lysates, tissue homogenates, serum, or plasma samples is incubated with cytokine ELISA plate, the captured cytokine proteins are subsequently detected with a cocktail of biotinylated detection antibodies. The test sample is allowed to react with pairs of two antibodies, resulting in the cytokines being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. The plate is further detected with HRP luminescent substrate. Luminescence is reported as relative light units (RLUs) on a microplate luminometer. The level of expression for each specific cytokine is directly proportional to the luminescent intensity.

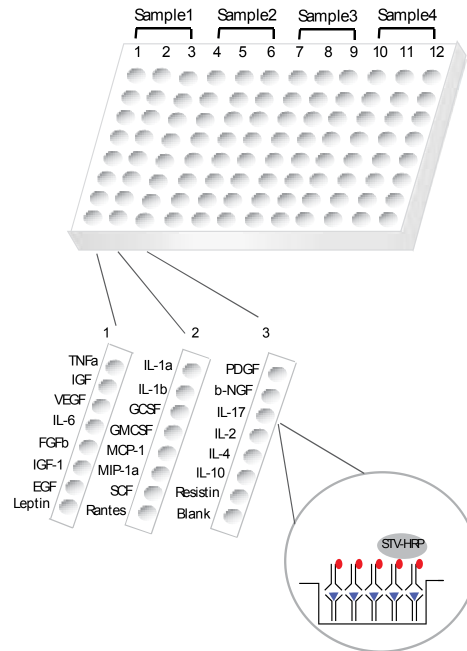

Diagram of mouse Cytokine ELISA plate array assay

Materials provided with the kit

- One white plate coated with 23 different antibodies against mouse cytokines (4°C).
- Biotin labeled antibody mixture against 23 different mouse cytokines (-20°C).
- Streptavidin-HRP conjugate (4°C)
- 1X Diluent buffer (4°C)
- 5X Assay wash buffer (RT)
- Substrate A (4°C)
- Substrate B (4°C)
- Substrate dilution buffer (RT)

Material required but not provided

- Luminometer plate reader
- Distilled H2O

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
  40ml 5x Assay wash buffer
  160ml ddH2O
- Sample preparation
  - Serum-free cell culture conditioned media can be used directly or dilute 2-fold with 1X Diluent buffer before use. When serum-containing conditioned media is required, be sure to use serum as a control.
  - Cell lysate or tissue lysate can be prepared following the protocol on our website: http://www.signosisinc.com/pdf/Preparation_of_Cell_Lysates_for_ELISA.pdf
  - Sera or plasma can be diluted 10-20 fold with 1X Diluent buffer.
- Dilute 400 times of biotin labeled antibody mixture with 1X Diluent buffer.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Cut the film over the plate and remove it from the desired number of well strips. Make sure the rest of wells are well sealed.
2. Prepare 3.5ml sample and add 100 µl per well to one section and incubate for 1 hour at room temperature with gentle shaking.
3. Invert the plate over an appropriate container and expel the contents forcibly. Wash the plate by adding 200µl of 1x Assay wash buffer. Repeat the washing process two times for a total of three washes. Complete removal of liquid at each wash by firmly tapping the plate against a pile of clean paper towels.
4. Add 100µl of diluted biotin-labeled antibody mixture to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100 µl of diluted streptavidin-HRP conjugate to each well and incubate for 30 min at room temperature with gentle shaking.
7. Repeat the aspiration/wash as in step 3.
8. Freshly prepare the substrate solution
   For the whole plate:
   1ml Substrate A
   1ml Substrate B
   8ml Substrate dilution buffer
9. Add 95µl substrate solution to each well and incubate for 2 minutes.
10. Place the plate in the luminometer, and read. Set integration time to 1 second with no filter position. For the best results, read the plate within 5-20 minutes.

Example of Analysis Data

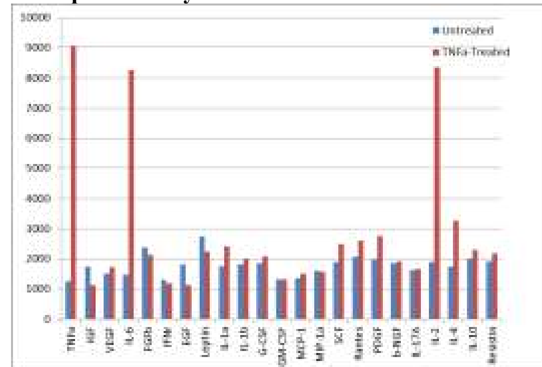

Figure1: Analysis of Cytokine Protein Expression in TNFα-Treated and Untreated HeLa with Mouse Cytokine ELISA Plate Array

NIH3T3 cells were starved for 24 hours with serum-free medium, subsequently treated the cells with and without 20ng/ul TNF for 16 hours. The serum-free conditioned media were incubated on the plate for 1 hour. After incubating with detection antibody mix and HRP, the plate was detected with chemilumincent substrate by a plate reader.

Diagram of Mouse Cytokine ELISA Plate Array I

	1	2	3	4	5	6	7	8	9	10	11	12
A	TNFa	IL-1a	PDGF	TNFa	IL-1a	PDGF	TNFa	IL-1a	PDGF	TNFa	IL-1a	PDGF
B	IGF	IL-1b	b-NGF	IGF	IL-1b	b-NGF	IGF	IL-1b	b-NGF	IGF	IL-1b	b-NGF
C	VEGF	G-CSF	IL-17A	VEGF	G-CSF	IL-17A	VEGF	G-CSF	IL-17A	VEGF	G-CSF	IL-17A
D	IL-6	GM-CSF	IL-2	IL-6	GM-CSF	IL-2	IL-6	GM-CSF	IL-2	IL-6	GM-CSF	IL-2
E	FGFb	MCP-1	IL-4	FGFb	MCP-1	IL-4	FGFb	MCP-1	IL-4	FGFb	MCP-1	IL-4
F	IFNr	MIP-1a	IL-10	IFNr	MIP-1a	IL-10	IFNr	MIP-1a	IL-10	IFNr	MIP-1a	IL-10
G	EGF	SCF	Resistin	EGF	SCF	Resistin	EGF	SCF	Resistin	EGF	SCF	Resistin
H	Leptin	Rantes	Blank	Leptin	Rantes	Blank	Leptin	Rantes	Blank	Leptin	Rantes	Blank