



Human MCP-1 ELISA

Catalog Number EA-0408

(For Research Use Only)

Introduction

Monocyte chemoattractant protein-1 (MCP-1), also called CCL2, is an inflammatory chemokine that plays important roles in recruiting monocytes, memory T cells, and dendritic cells to sites of tissue injury and infection. MCP-1 also involves in obesity and insulin resistance by the induction of an inflammatory response (macrophage infiltration) in fatty tissue. In addition, MCP-1 has been found in the joints of people with rheumatoid arthritis where may serve to recruit macrophages and perpetuate the inflammation in the joints.

Principle of the assay

MCP-1 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-human MCP-1 antibodies for immobilization on the microtiter wells and rabbit anti-human MCP-1 antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the MCP-1 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of MCP-1 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

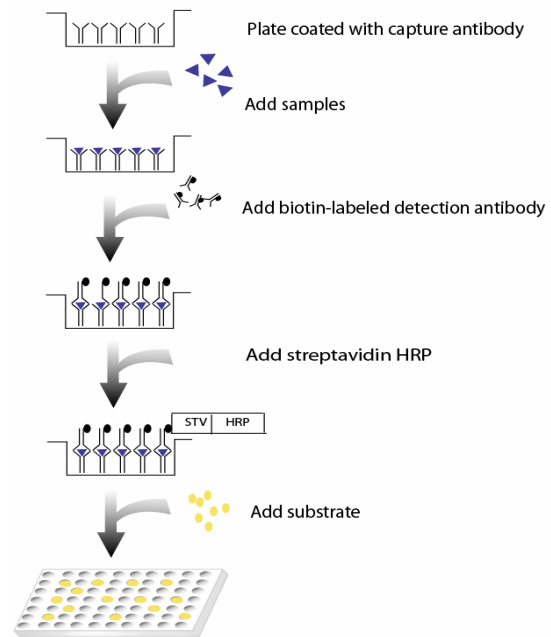


Diagram of ELISA

Materials provided with the kit

- 8x12 96-well microplate coated with rabbit anti-human MCP-1 antibodies (4°C).
- Biotin labeled rabbit anti-human MCP-1 antibody (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Recombinant human MCP-1 standard (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution (4°C).

Material required but not provided

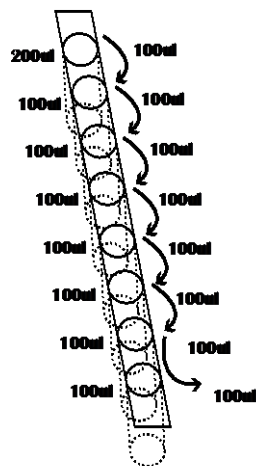
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
40ml 5x Assay wash buffer
160ml ddH₂O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 50 times of human recombinant MCP-1 (400ng/ml) with 1X Diluent buffer to 8000pg/ml and then 2-fold serial dilutions. To dilute 50 times of Human MCP-1, add 4ul Human Recombinant MCP-1 in 200ul 1X Diluent Buffer (See Step 2 in "Assay Procedure" for detailed instruction)
- Dilute 400 times of biotin labeled rabbit anti-human MCP-1 with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Calculate the number of samples to decide how many strips need to be used.
2. Add 100 μ l of samples or standard to each well. See instruction and diagram below for standard preparation.



- a. Add 200ul 1X Diluent buffer to the 1st well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
- b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
- c. Mix dilutions in 1st well and transfer 100ul from the 1st well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Aspirate each well and wash by adding 200 μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
4. Add 100 μ l of diluted biotin-labeled goat anti-human MCP-1 antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100 μ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.

7. Repeat the aspiration/wash as in step 3.
8. Add 100 μ l substrate to each well and incubate for 10-30 minutes.
9. Add 50 μ l of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.