



## Rat IP-10 ELISA

Catalog Number EA-3012

(For Research Use Only)

### Introduction

IFN-gamma-inducible protein (IP-10) is a member of the chemokine family of cytokines and is induced in a variety of cells in response to interferon gamma and lipopolysaccharide. It is secreted by a number of cells including monocytes, endothelial cells and fibroblasts. IP-10 plays several roles, such as chemoattraction for monocytes/macrophages, T cells, NK cells, and dendritic cells, promotion of T cell adhesion to endothelial cells, antitumor activity, and inhibition of bone marrow colony formation. Several cell types in response to IFN- $\gamma$ . IP-10 acts as potent inhibitors of angiogenesis in vivo.

### Principle of the assay

IP-10 ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-rat IP-10 for immobilization on the microtiter wells and biotinylated rabbit anti-rat IP-10 antibody along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IP-10 molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IP-10 is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Diagram of ELISA

### Materials provided with the kit

Component	Qty	Store at
8x12 96-well microplate coated with rabbit anti-rat IP-10 antibody	1	4°C
Biotin labeled rabbit anti-rat IP-10 antibody	25 $\mu$ L	-20°C
Recombinant rat IP-10 standard (400ng/ml)	5 $\mu$ L	-20°C
Streptavidin-HRP conjugate	50 $\mu$ L	4°C
1xDiluent buffer	40mL	4°C
5X Assay wash buffer	40mL	4°C
Substrate	10mL	4°C
Stop solution	5mL	4°C

### Material required but not provided

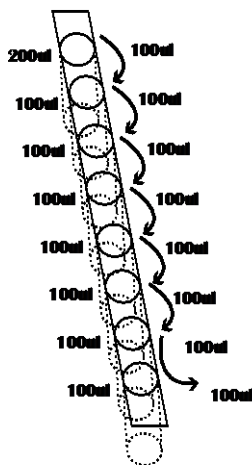
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer  
40ml 5x Assay wash buffer  
160ml ddH<sub>2</sub>O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 100 times of rat recombinant IP-10 (400ng/ml) to 4ng/ml by adding 2ul rat IP-10 recombinant in first well with 200ul of 1X Diluent buffer and then 2-fold serial dilutions.
- Dilute 400 times of biotin labeled rabbit anti-rat IP-10 antibody with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

## Assay procedure

1. Take the desired the number of samples to decide how many strips need to be used. Make sure the rest of strip are well sealed.
2. Standard curve:



- a. Add 200ul 1X Diluent buffer to the 1<sup>st</sup> well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
- b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
- c. Mix dilutions in 1<sup>st</sup> well and transfer 100ul from the 1<sup>st</sup> well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Add 100ul of sample per well and incubate for 1 hour at room temperature with gentle shaking.
4. Aspirate each well and wash by adding 200ul of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100ul of diluted biotin-labeled anti-rat IP-10 to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4

7. Repeat the aspiration/wash as in step 4.
8. Add 100ul of substrate to each well and incubate for 5-30 minutes.
9. Add 50ul of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.