Signosis
Innovative Plate Assay Solutions

Human SCF ELISA

Catalog Number EA-0407 **(For Research Use Only)**

Introduction

Stem cell factor (SCF) is a growth factor for multiple cell types. It is expressed in glioma cells and as a result of various types of brain injury. Tumor-induced brain injury, brain cell-mediated SCF expression contributes to tumor growth by setting up an environment that supports angiogenesis and tumor progression. SCF expression is not directly linked to tumor cell proliferation but instead encourages the growth of blood vessels needed to support the expanding tumor.

Principle of the assay

SCF ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-human SCF antibodies for immobilization on the microtiter wells and rabbit anti-human SCF antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the SCF molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of SCF is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.

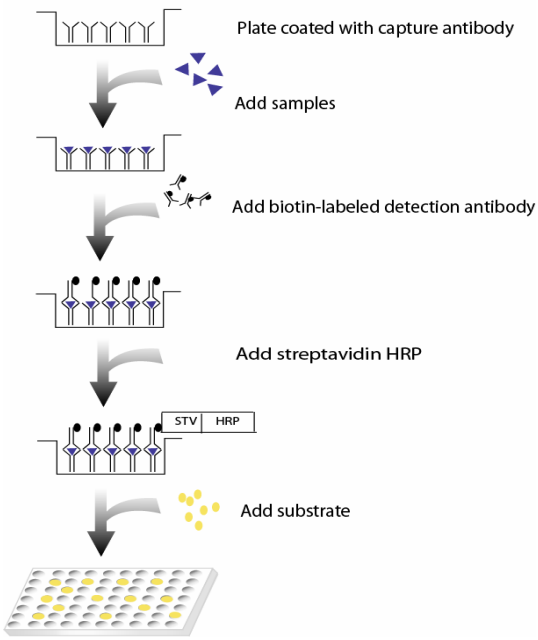


Diagram of ELISA

Materials provided with the kit

- 8x12 96-well microplate coated with rabbit anti-human SCF antibodies (4ºC).
- Biotin labeled rabbit anti-human SCF antibodies (-20ºC).
- Streptavidin-HRP conjugate (4ºC).
- Recombinant human SCF standard (1000ng/ml) (-20ºC).
- 1X Diluent buffer (4ºC).
- 5X Assay wash buffer (RT)
- Substrate (4ºC).
- Stop Solution (4ºC).

Material required but not provided

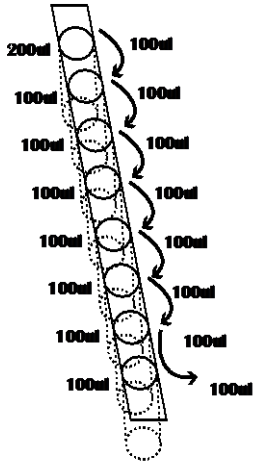
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer
  40ml 5x Assay wash buffer
  160ml ddH2O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 100 times of human recombinant SCF (400ng/ml) with 1X Diluent buffer to 4000pg/ml and then 2-fold serial dilutions by adding 2µl human recombinant SCF in 200ul 1x Diluent Buffer (see step2 below for the detailed instruction).
- Dilute 400 times of biotin labeled rabbit anti-human SCF antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

Assay procedure

1. Calculate the number of samples to decide how many strips need to be used. Make sure the rest wells are well sealed.
2. Add 100µl of Standard, control, or sample per well and incubate for 1 hour at room temperature with gentle shaking.



a. Add 200ul 1X Diluent buffer to the 1st well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
c. Mix dilutions in 1st well and transfer 100ul from the 1st well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Aspirate each well and wash by adding 200µl of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
4. Add 100µl of diluted biotin-labeled rabbit anti-human SCF antibodies to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100 µl of diluted streptavidin-HRP conjugate to
7. Repeat the aspiration/wash as in step 3.
8. Add 100µl substrate to each well and incubate for 5-30 minutes.
9. Add 50µl of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

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