



## Mouse EGF ELISA

Catalog Number EA-2403

(For Research Use Only)

### Introduction

Epidermal growth factor (EGF) is a strong mitogen for many cells, such as fibroblasts. EGF also stimulates the proliferation of embryonic cells. The proliferation of some cell lines has been shown to be inhibited by EGF. EGF acts as a differentiation factor for some cell types. To a limited extent EGF also augments angiogenesis because it is mitogenic for endothelial cells. The mitogenic activity of EGF for endothelial cells can be potentiated by thrombin. EGF is a strong chemoattractant for fibroblasts and epithelial cells. EGF alone and also in combination with other cytokines is an important factor mediating wound healing processes.

### Principle of the assay

EGF ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes goat anti-mouse EGF antibodies for immobilization on the microtiter wells and goat anti-mouse EGF antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the EGF molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of EGF is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Diagram of ELISA

### Materials provided with the kit

- 8x12 96-well microplate coated with goat anti-mouse EGF antibodies (4°C).
- Biotin labeled goat anti-mouse EGF antibodies (-20°C).
- Streptavidin-HRP conjugate (4°C).
- Mouse recombinant EGF standard (1000ng/ml) (-20°C).
- 1X Diluent buffer (4°C).
- 5X Assay wash buffer (RT)
- Substrate (4°C).
- Stop Solution (4°C).

### Material required but not provided

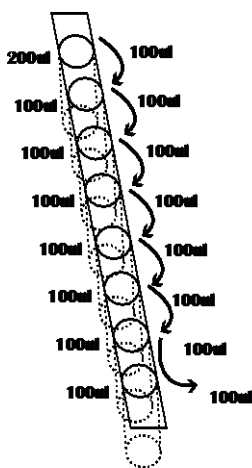
- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

## Reagent preparation before starting experiment

- Dilute the 5x Assay wash buffer to 1x buffer  
40ml 5x Assay wash buffer  
160ml ddH<sub>2</sub>O
- Use serum-free conditioned media or original or 10-fold diluted sera. Sera can be diluted with 1 X Diluent buffer. When serum-containing conditioned media is required, be sure to use serum as a control.
- Dilute 500 times of mouse recombinant EGF (1000ng/ml) with 1X Diluent buffer to 2000pg/ml and then 2-fold serial dilutions.
- Dilute 400 times of biotin labeled goat anti-mouse EGF antibodies with 1X Diluent buffer before use.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer before use.

## Assay procedure

1. Calculate the number of samples to decide how many strips need to be used.
2. Add 100  $\mu$ l of Standard, control, or sample per well and incubate for 1 hour at room temperature with gentle shaking.



- a. Add 200ul 1X Diluent buffer to the 1<sup>st</sup> well. Add 100ul 1X Diluent Buffer to the rest wells of strip.
- b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
- c. Mix dilutions in 1<sup>st</sup> well and transfer 100ul from the 1<sup>st</sup> well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Aspirate each well and wash by adding 200ul of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Complete removal of liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
4. Add 100ul of diluted biotin-labeled goat anti-mouse EGF antibodies to each well and incubate for 1 hour at room temperature with gentle shaking.
5. Repeat the aspiration/wash as in step 3.
6. Add 100  $\mu$ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.

7. Repeat the aspiration/wash as in step 3.
8. Add 100ul substrate to each well and incubate for 5-30 minutes.
9. Add 50ul of Stop solution to each well. The color in the wells should change from blue to yellow.
10. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.

## Example of standard curve

