



Mouse IL-1 β ELISA

Catalog Number EA-2508

(For Research Use Only)

Introduction

IL-1 α and IL-1 β are prominent agonists mediating inflammatory and immunomodulatory effects. Both are produced by macrophages, monocytes and dendritic cells. They are important part of the inflammatory response against infection. They increase the expression of adhesion factors on endothelial cells to enable transmigration of leukocytes to sites of infection. IL-1 β production in peripheral tissue has also been associated with hyperalgesia (increased sensitivity to pain) associated with fever.

Principle of the assay

IL-1 β ELISA is based on the principle of a solid phase enzyme-linked immunosorbent assay. The assay utilizes rabbit anti-mouse IL-1 β for immobilization on the microtiter wells and biotinylated rabbit anti-mouse IL-1 β antibodies along with streptavidin conjugated to horseradish peroxidase (HRP) for detection. The test sample is allowed to react simultaneously with the two antibodies, resulting in the IL-1 β molecules being sandwiched between the solid phase and enzyme-linked antibodies. After incubation, the wells are washed to remove unbound-labeled antibodies. A HRP substrate, TMB, is added to result in the development of a blue color. The color development is then stopped with the addition of Stop Solution changing the color to yellow. The concentration of IL-1 β is directly proportional to the color intensity of the test sample. Absorbance is measured spectrophotometrically at 450 nm.



Diagram of ELISA

Materials provided with the kit

Component	Qty	Store at
8x12 96-Well 12 strip Plate coated with rabbit anti-mouse IL-1β antibodies	1	4°C
Biotin labeled rabbit anti-mouse IL-1β antibodies	25 μ L	-20°C
Recombinant mouse IL-1β standard (200ng/ml)	10 μ L	-20°C
Streptavidin-HRP conjugate	50 μ L	4°C
1xDiluent buffer	40mL	4°C
5X Assay wash buffer	40mL	4°C
Substrate	10mL	4°C
Stop solution	5mL	4°C

Material required but not provided

- Microplate reader capable of measuring absorbance at 450 nm
- Deionized or distilled water.

Reagent preparation before starting experiment

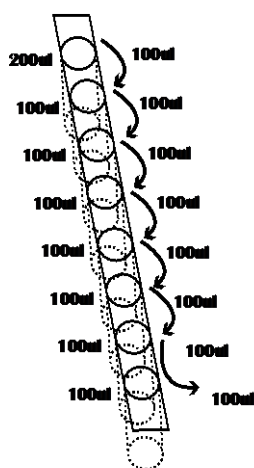
- Dilute the 5x Assay wash buffer to 1x buffer
40ml 5x Assay wash buffer
160ml ddH₂O
- Dilute 50 times of mouse IL-1 β recombinant protein (400ng/ml) with 1X Diluent buffer to 4ng/ml by adding 4 μ l Mouse IL-1 β recombinant protein in first well with 200 μ l diluent buffer then 2-fold serial dilutions (see step 2 in Assay procedure below for the detailed instruction).
- Dilute 400 times of biotin labeled mouse anti IL-1 β antibody with 1X Diluent buffer.
- Dilute 200 times of streptavidin-HRP with 1X Diluent buffer.

Sample preparation before starting experiment

- For **media samples**, add 100 μ l directly to the well.
- For **cell lysate samples**, use cell lysis buffer (Catalog# EA-0001). Follow protocol in Cell Lysate Buffer User Manual.
- For **serum or plasma samples**, we recommend a 1:10 dilution with 1X diluent buffer, for example, add 10 μ l sample in 90 μ l 1X diluent buffer.

Assay procedure

1. Take the desired the number of samples to decide how many strips need to be used. Make sure the rest of strips are well sealed.
2. Standard curve:



- a. Add 200 μ l 1X Diluent buffer to the 1st well. Add 100 μ l 1X Diluent Buffer to the rest wells of strip.
- b. Add appropriate amount of protein recombinant (follow instruction in "Reagent Preparation")
- c. Mix dilutions in 1st well and transfer 100 μ l from the 1st well to the next dilution. (See picture) Incubate each well for 1 hr at room temperature with gentle shaking

3. Add 100 μ l of sample to each well, incubate for 1-2 hours at room temperature with gentle shaking

4. Aspirate each well and wash by adding 200 μ l of 1X Assay wash buffer. Repeat the process three times for a total of three washes. Completely remove liquid at each wash. After the last wash, remove any remaining liquid by inverting the plate against clean paper towels.
5. Add 100 μ l of diluted biotin-labeled anti-mouse IL-1 β antibody to each well and incubate for 1 hour at room temperature with gentle shaking.
6. Repeat the aspiration/wash as in step 4.
7. Add 100 μ l of diluted streptavidin-HRP conjugate to each well and incubate for 45 min at room temperature with gentle shaking.
8. Repeat the aspiration/wash as in step 4.
9. Add 100 μ l of substrate to each well and incubate for 5-30 minutes.
10. Add 50 μ l of Stop solution to each well. The color in the wells should change from blue to yellow.
11. Determine the optical density of each well with a microplate reader at 450 nm within 30 minutes.