



## pMiR-Luc Reporter Vectors

Catalog Number LR-XXXX

(For Research Use Only)

### Introduction

pMiR-Luc<sup>TM</sup> reporter vectors are a series of firefly luciferase-based reporter constructs for quantitative measurement of miRNA expression in cells. Each vector contains the CMV promoter, firefly luciferase gene, a unique miRNA target site at 3'UTR, and a SV40 terminator sequence. The target site is a sequence perfectly complementary to a specific miRNA. When the miRNA is expressed, it binds to the sequence and results in repression of luciferase gene expression. Therefore, luciferase activity represents the expression and activity of a miRNA. When pMiR-Luc reporter vectors are transfected into mammalian cells, they can be used to detect endogenous miRNA expression and activity, or used to monitor the up- or down-regulation of miRNAs.

### Recommend transfection and assay

We recommend using FuGENE<sup>TM</sup> 6 (Roche) for the transfection of pMiR-Luc reporter vectors, as the use of other transfection methods could lead to reduced luciferase activity from the reporters.

1. Plate  $1-3 \times 10^5$  cells in 1 ml of growth medium containing serum without antibiotics in a 12-well culture plate at one day before transfection, which will yield 50-80% confluence on the day of transfection. We recommend to plate cells in duplicate.
2. For each transfection, dilute 0.2  $\mu\text{g}$  of the reporter vector with 50  $\mu\text{l}$  of Opti-MEM I Reduced Serum Medium or serum-free culture media and dilute 3  $\mu\text{l}$  FuGENE 6 Reagent with another 50  $\mu\text{l}$  of Opti-MEM I Reduced Serum Medium or serum-free culture media, mix, and incubate for 5 min at room temperature, but no longer. Combine the diluted mix. Incubate for 15-30 min at room temperature. Once the FuGENE 6 Reagent is diluted, it needs to use within 45 min.
3. Add 100  $\mu\text{l}$  of DNA/FuGENE mix to the complete growth media on cells and mix gently by rocking the plate back and forth. Incubate the cells at 37°C in a CO<sub>2</sub> incubator, overnight.
4. Measure luciferase expression 24-48 hr after transfection. Aspirate to completely remove the media from the culture plates.

5. Lyse the attached cells by adding lysis buffer to each well. Use approximately 50  $\mu\text{l}$  per well for a 12-well plate. To detach cells from the plate, pipet the mixture up and down. Transfer the cell lysate/buffer solution to a clean 1.5-ml microcentrifuge tube. Keep on ice or store at -20°C. Assay for luciferase activity following the instructions given by the supplier.

### E. coli transform to propagate the plasmids

1. Transform *E. coli* competent cells with the plasmid.
2. Plate the transformed cells on LB plates containing 100  $\mu\text{g}/\text{mL}$  Ampicillin and grow overnight at 37°C.
3. Transfer a single colony to 1-2 ml LB medium containing 100 $\mu\text{g}/\text{mL}$  and shake at 37°C overnight.
4. Prepare plasmids and check on gel.

### Diagram of pMiR-Luc reporter vectors

